

## Daphniglaucin C, a novel tetracyclic alkaloid from *Daphniphyllum glaucescens*

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**Abstract**—A novel *Daphniphyllum* alkaloid with an unprecedented tetracyclic ring system consisting of an octahydroindole and a hexahydroazulene rings, daphniglaucin C (**1**), has been isolated from the leaves of *Daphniphyllum glaucescens* and the structure and relative stereochemistry were elucidated on the basis of spectroscopic data. Daphniglaucin C (**1**) inhibited the polymerization of tubulin.

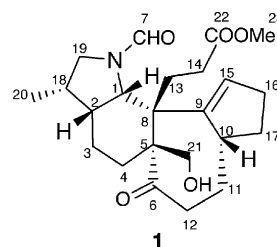
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Plants of *Daphniphyllum* species produce structurally diverse alkaloids with unusual polycyclic skeletons.<sup>1,2</sup> These unique ring systems have attracted great interest as challenging targets for total synthesis and biosynthetic studies.<sup>3</sup> Heathcock and co-workers have proposed a biogenetic pathway for *Daphniphyllum* alkaloids and demonstrated a biomimetic total synthesis of several *Daphniphyllum* alkaloids.<sup>3,4</sup>

Recently, we have isolated some novel types of *Daphniphyllum* alkaloids<sup>5–12</sup> such as daphnezomines A and B<sup>5</sup> with a unique aza-adamantane core and daphnezomines F and G<sup>6</sup> with a 1-azabicyclo[5.2.2]undecane ring system as well as daphnicyclidins A–H,<sup>8</sup> J, and K<sup>9</sup> with unique hexa- or pentacyclic ring system, and daphmanidin A<sup>10</sup> with an unprecedented fused-hexacyclic skeleton from the leaves and stems of *D. teijsmanni* and/or *D. humile*, daphniglaucin A<sup>11</sup> with a fused-heptacyclic skeleton and a quaternary nitrogen from the leaves of *D. glaucescens*, and calyciphyllines A and B<sup>12</sup> with a novel hexacyclic skeleton from the leaves of *D. calycinum*. In our continuing search for structurally unique and biogenetically interesting *Daphniphyllum* alkaloids, daphniglaucin C (**1**), a novel tetracyclic alkaloid consisting of an octahydroindole and a hexahydroazulene

rings, was isolated from the leaves of *D. glaucescens*. This paper describes the isolation and structural elucidation of **1**.

The leaves of *D. glaucescens* were extracted with MeOH, and the extract was partitioned between EtOAc and 3% tartaric acid. Water-soluble materials, which were adjusted at pH 9 with satd Na<sub>2</sub>CO<sub>3</sub>, were extracted with CHCl<sub>3</sub>. CHCl<sub>3</sub>-soluble materials were subjected to an amino silica gel column (hexane/EtOAc, 9:1 → 1:1 and then CHCl<sub>3</sub>/MeOH, 1:0 → 0:1), from which a fraction eluted with CHCl<sub>3</sub>/MeOH (7:3) was purified by C<sub>18</sub> HPLC (30% CH<sub>3</sub>CN/0.1% TFA) to afford daphniglaucin C<sup>13</sup> (**1**, 0.009% yield) together with a known alkaloid, macrodaphniphyllidine.<sup>14</sup>



**Keywords:** Alkaloid; *Daphniphyllum glaucescens*; Daphniglaucin C; NMR.

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Daphniglaucin C (**1**) showed the pseudomolecular ion peak at  $m/z$  404 ( $M+H$ )<sup>+</sup> in the FABMS, and the molecular formula, C<sub>23</sub>H<sub>33</sub>NO<sub>5</sub>, was established by HRFABMS [ $m/z$  404.2438, ( $M+H$ )<sup>+</sup>,  $\Delta$  +0.1 mmu]. IR

absorptions implied the presence of hydroxyl ( $3370\text{ cm}^{-1}$ ) and three carbonyl functionalities including ester, ketone, and amide ( $1730$ ,  $1695$ , and  $1635\text{ cm}^{-1}$ , respectively).  $^{13}\text{C}$  NMR data (Table 1) revealed 23 carbon signals due to one trisubstituted olefin, one carbonyl, one ester carbonyl, one amide carbonyl, two  $\text{sp}^3$  quaternary carbons, four  $\text{sp}^3$  methines, ten  $\text{sp}^3$  methylenes, one methyl, and one methoxy group. Among them, one methylenes ( $\delta_{\text{C}} 51.7$ ;  $\delta_{\text{H}} 2.96$  and  $3.61$ ) and one methine ( $\delta_{\text{C}} 66.3$ ;  $\delta_{\text{H}} 4.15$ ) were ascribed to those bearing a nitrogen, while one methylene ( $\delta_{\text{C}} 65.4$ ;  $\delta_{\text{H}} 3.81$  and  $4.49$ ) was that bearing an oxygen.

The  $^1\text{H}$ - $^1\text{H}$  COSY and HOHAHA spectra revealed connectivities of three partial structures **a** (C-1 to C-4, C-2 to C-18, and C-18 to C-19 and C-20), **b** (C-10 to C-12, C-10 to C-17, and C-15 to C-17), and **c** (C-13 to C-14) as shown in Figure 1. HMBC correlations were observed for H-19b to C-1 ( $\delta_{\text{C}} 66.3$ ) and H-7 to C-19 ( $\delta_{\text{C}} 51.7$ ), suggesting that C-1, C-7, and C-19 were connected to each other through a nitrogen atom. Long-range couplings for H-7 to H<sub>2</sub>-19 and H-1 indicated the presence of an *N*-formyl group ( $\delta_{\text{H}} 8.08$ ;  $\delta_{\text{C}} 165.3$ ). The connectivity of C-1 and C-13 through C-8 was implied by HMBC correlations for H-1 and H-13 to C-8 ( $\delta_{\text{C}} 46.8$ ). HMBC correlations for H-4 to C-5 ( $\delta_{\text{C}} 57.6$ ) and H<sub>2</sub>-21 to C-5 and C-6 ( $\delta_{\text{C}} 216.0$ ) indicated that C-21 was connected to C-4 and C-6 through C-5. HMBC cross-

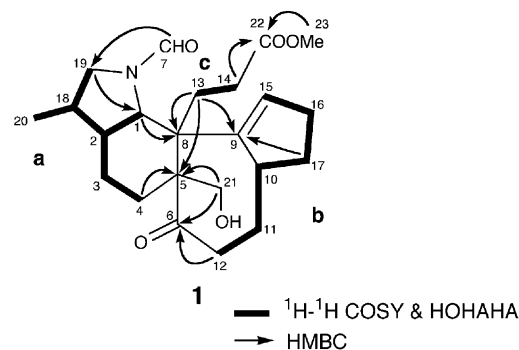


Figure 1. Selected 2D NMR correlations for daphniglaucin C (**1**).

peaks for H<sub>2</sub>-13 to C-5 and C-9 ( $\delta_{\text{C}} 150.6$ ) indicated connectivities of C-8 to C-5 and C-9, constructing an octahydroindole ring system. The connectivity of C-6 to C-12 was implied by the HMBC correlation for H-12b to C-6. In addition, the HMBC correlation for H-17 to C-9 indicated the connectivity of C-9 to C-10, constructing a hexahydroazulene ring system. A methoxy group was attached to C-22 by HMBC correlations for H<sub>3</sub>-23 and H<sub>2</sub>-14 to C-22 ( $\delta_{\text{C}} 175.6$ ). Thus, the gross structure of daphniglaucin C was assigned as **1** having an unprecedented fused-tetracyclic ring system consisting of an octahydroindole ring with an *N*-formyl group and a methyl group (C-20) at C-18 and a hexahydroazulene ring with a ketone group at C-6 and a methoxycarbonyl ethyl group (C-13, C-14, C-22, and C-23) at C-8 as shown in Figure 1.

Table 1.  $^1\text{H}$  [ $\delta_{\text{H}}$  (*J*, Hz)] and  $^{13}\text{C}$  [ $\delta_{\text{C}}$ ] NMR data of daphniglaucin C (**1**) in  $\text{CD}_3\text{OD}$  at 300 K

	$\delta_{\text{H}}$	$\delta_{\text{C}}$	HMBC ( $^1\text{H}$ )
1	4.15 (1H, d, 4.1)	66.3	13, 19b
2	2.24 (1H, m)	40.4	18, 19b, 20
3a	1.65 (1H, m)	16.9	—
3b	1.76 (1H, m)	—	—
4a	1.60 (1H, m)	26.7	21
4b	2.22 (1H, m)	—	—
5	—	57.6	4b, 13, 21
6	—	216.0	12b, 21
7	8.08 (1H, s)	165.3	—
8	—	46.8	1, 13
9	—	150.6	13, 15, 16a, 17a
10	3.16 (1H, br t, 11.5)	47.1	12a, 15, 17b
11a	1.55 (1H, m)	32.3	17b
11b	1.88 (1H, m)	—	—
12a	2.32 (1H, m)	42.9	—
12b	3.20 (1H, dt, 5.9, 13.2)	—	—
13	2.01 (2H, t, 7.8)	31.3	14
14	2.25 (2H, t, 7.8)	30.6	13
15	5.74 (1H, s)	132.0	16a, 17a
16a	2.44 (1H, m)	30.1	15, 17b
16b	2.30 (1H, m)	—	—
17a	1.74 (1H, m)	34.5	15, 16b
17b	2.16 (1H, m)	—	—
18	2.29 (1H, m)	34.9	19a, 20
19a	3.61 (1H, t, 11.5)	51.7	7, 20
19b	2.96 (1H, t, 11.5)	—	—
20	1.09 (3H, d, 6.7)	12.8	19a
21	4.49 (1H, d, 10.6)	65.4	—
	3.81 (1H, d, 10.6)	—	—
22	—	175.6	13, 14, 23
23	3.65 (3H, s)	52.3	—

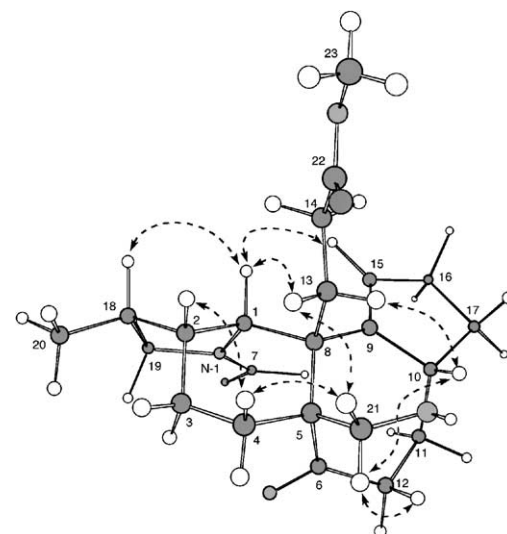
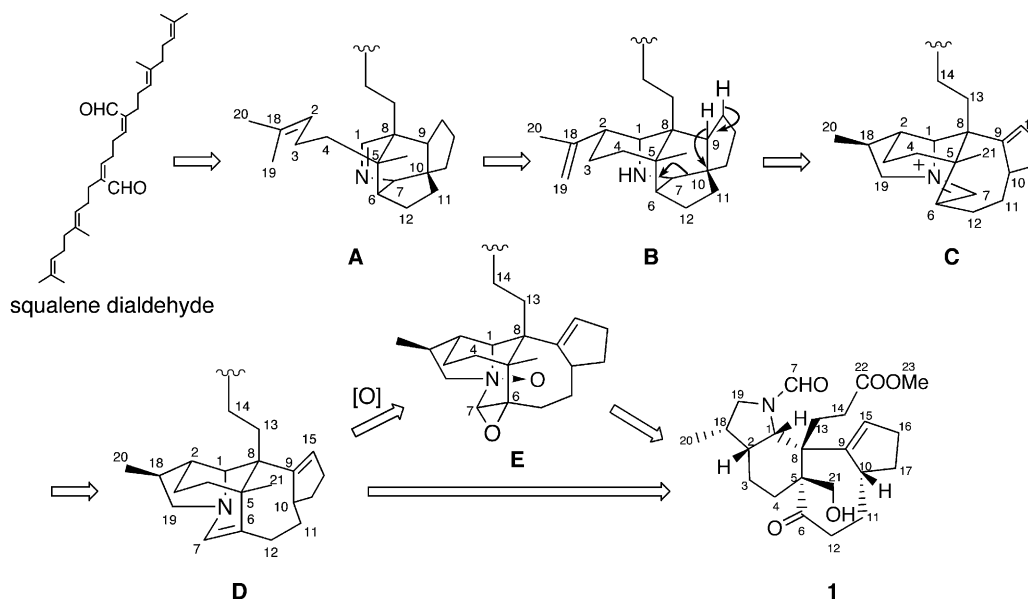


Figure 2. Selected NOESY correlations (dotted arrows) and relative stereochemistry for daphniglaucin C (**1**).



Scheme 1. Plausible biogenetic path for daphniglaucin C (1).

A plausible biogenetic pathway for daphniglaucin C (1) is proposed as shown in Scheme 1. The biogenetic origin of daphniglaucin C (1) seems to be an imine intermediate C, which might be produced through fragmentation reaction of a secodaphnane skeleton (B) derived from an imine intermediate A proposed by Heathcock et al.<sup>3</sup> Oxidation of N-1, C-6, and C-7 of the intermediate D and cleavage of C-6 to C-7 bond of an intermediate E by Polonovski-type reaction<sup>15</sup> will give the skeleton of daphniglaucin C (1), although an alternative path through oxidative cleavage of C-6 to C-7 bond is also possible.

Daphniglaucin C (1) exhibited cytotoxicity against murine lymphoma L1210 cells ( $IC_{50}$ , 0.1  $\mu\text{g/mL}$ ) in vitro and inhibited the polymerization of tubulin<sup>16,17</sup> ( $IC_{50}$ , 25  $\mu\text{M}$ ).

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